SESOUITERPENOIDS OF RICCARDIA AND PALLAVICINIA SPECIES

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(Revised received 14 July 1980)

Key Word Index—Aneura pinguis; Riccardia multifida; R. jackii; Riccardiaceae; Pallavicinia longispina; Dilaenaceae; Metzgeriales; Hepaticae; ent-sesquiterpenes; ent-selinanes; ent-bicyclogermacrane; ent-aromadendrane; (R)-cuprane; chemosystematics.

Abstract—Riccardia species (Metzgeriales) contain various types of sesquiterpenes. R. jackii produces ent-selinane-, ent-aromadendrane- and ent-bicyclogermacrane-type sesquiterpenes together with (R)-cuparene and α -barbatene. Aneura pinguis (= Riccardia pinguis) is chemically quite different from R. multifida and R. jackii. The former produces a large amount of pinguisone. R. multifida contains 6-(3-methyl-2-butenyl)-indole and (+)- β -elemene as the major components. Pallavicinia longispina (Dilaenaceae; Metzgeriales) produces mainly spathulenol. The chiral properties of the sesquiterpenes isolated from R. jackii are quite similar to those of red algae, Laurencia species.

INTRODUCTION

The results of the chemical analyses of ninety-eight species of Japanese and European liverworts have shown that members of the Hepaticae contain compounds which are of chemosystematic value [1-6]. The presence of carotenoids [7], lower carboxylic acids [8], carbohydrates [9, 10], indole derivatives [11, 12] and bibenzyl derivatives [13] has been reported for a few Riccardia species (Riccardiaceae; Metzgeriales). Recently, Benesova et al. [14] reported on the isolation of a unique sesquiterpene, pinguisone (7), from Aneura pinguis Dum. (=Riccardia pinguis). Except for the isolation of pinguisone, there are few reports concerning the presence of terpenoids in *Riccardia* species [15]. In Japan, five genera, Makinoa, Pellia, Calycularia, Pallavicinia and Moerckia constitute the Dilaenaceae, which is situated systematically near the Riccardiaceae. Recently, we reported that Makinoa crispata (Steph.) Miyake and Pellia endiviifolia (Dicks.) Dum. produce sesquiterpene lactones and diterpene dialdehydes as the major components, respectively [1, 16, 17]. In the present communication, we wish to report the distribution of sesquiterpenoids in three Riccardia species and Pallavicinia longispina Steph.

RESULTS AND DISCUSSION

Air-dried ground materials were extracted with Et₂O. Each crude extract was analysed by GC-MS to obtain an indication of the compounds present. The chemical structures of the terpenoids were confirmed by comparison of the MS spectra with those of authentic samples or with published data. The major components, which appeared to be of chemotaxonomic value were isolated by PLC or preparative GLC.

Riccardia multifida S. Gray contains a large amount of 6-[3-methyl-2-butenyl]-indole (8). This compound has already been found in European R. incurvata Lindb. and R. sinuata (With.) Grolle (=R. chamedryfolia) [11, 12].

Thus, Japanese R. multifida is chemically close to these two European species. In addition to the indole derivative, R. multifida contains the sesquiterpene hydrocarbon (+)- β -elemene (1). α -Elemene, γ -cadinene and calamenene were detected by GC-MS analysis.

A. pinguis (=R. pinguis) is chemically very specific, since it produces mainly a unique furanosesquiterpene, pinguisone (7), which has also been isolated from the same European species [14]. Recently, 7 and its related furanosesquiterpenes have been found in Jungermanniales: Porella, Lejeunea, Ptilidium and Trichocoleopsis [5, 18-22], although they are morphologically quite different from Riccardia.

Riccardia jackii Schiffn. produces various types of sesquiterpenoids, particularly hydrocarbons, e.g. $(+)-\alpha$ -selinene (2), $(-)-\beta$ -selinene (3), (-)-bicyclogermacrene (4), (+)-cuparene (5) and $(+)-\alpha$ -barbatene (9) and alcohols, e.g. (-)-spathulenol (6). All these sesquiterpenes have been found in various species of Jungermanniales. R. jackii and R. multifida are morphologically different from A. pinguis. From the sesquiterpene constitution described above, it is obvious that A. pinguis is also chemically very different from R. multifida and R. jackii.

Makinoa crispata, Pellia endiviifolia and Pallavicinia longispina are morphologically similar and they are placed in the Dilaenaceae which is close to the Riccardiaceae. M. crispata chemically resembles P. endiviifolia since they both contain unique sacculatane-type diterpene dialdehydes [1, 16, 17]. P. longispina morphologically resembles Pellia species rather than M. crispata. It is clear that P. longispina is chemically quite different from Makinoa and Pellia species, since it elaborates a large amount of ent-spathulenol (6) together with much triglycerides in place of diterpene dialdehydes.

Most liverworts biosynthesize the enantiomers of the sesquiterpenoids found in higher plants, although there are several exceptions. The sesquiterpenes found in R. jackii are interesting from both a biogenetic and an evolutionary view point. The selinenes (2 and 3),

spathulenol (6) and bicyclogermacrene (4), which may be a precursor of 6, are the enantiomers of the corresponding sesquiterpenes found in higher plants. The configuration of the cuparene series isolated from the liverworts so far examined is opposite to that isolated from the higher plants [23, 24]. In contrast, cuparene (5) isolated from R. jackii has the same configuration as that found in higher plants. Recently, many marine plants have been chemically investigated and various types of sesqui- and di-terpenoids have been isolated and their chemical

structures established. As seen in Fig. 1, co-occurrence of ent-elemene (1), ent-selinenes (2, 3) and (R)-(+)-cuparene (5) in Riccardia species has also been found in red algae, Laurencia species [25-27]. Roughly speaking, the Hepaticae are chemically quite different from the Musci. The former are similar to the higher plants and algae, whilst the latter resemble the pteridophytes [1, 3], although there is a large evolutionary distance between them. The morphological similarity of Riccardia species to some red algae and the chiral properties of the

Hepaticae Riccardia	Algae Laurencia	Higher plants
	₩ H	H
H 2	HO H CI	H
	Br Cl	H
4		
5*		
HO H		но

Fig. 1. Chiral properties of sesquiterpenoids among Hepaticae, algae and higher plants.* (S)-(-)-Cuparene has also been isolated from Jungermanniales.

sesquiterpenoids isolated from R. jackii led us to put forward the hypothesis that there is an evolutionary relationship between the Hepaticae and algae.

EXPERIMENTAL

 $^1\text{H NMR}$: 60 MHz in CDCl₃; IR: film: MS (direct inlet system or GC–MS): 70 eV; Optical rotations: CHCl₃. GC–MS: glass column, SE-30 1%; 50–270°, 5°/min; He 30 ml/min. Preparative GLC: SE-30 10%, 3 m \times 2 mm glass column; 80°–240°, 5°/min; N $_2$ 30 ml/min. TLC: precoated Si gel (0.25 mesh) F $_{254}$, n-hexane–EtOAc (4:1) and C $_6\text{H}_6$ –EtOAc (4:1). Spots were detected by spraying 30 % H $_2\text{SO}_4$ and heating at 100°, and by UV light (254 nm).

Plant material. The species, identified by Drs. S. Hattori and M. Mizutani, are deposited in the Herbarium, Hattori Botanical Laboratory and Tokushima Bunri University, Inst. of Pharmacognosy.

Extraction and isolation. R. multifida S. Gray collected in Gotaki, Tokushima prefecture in May 1979 was air-dried for 2 days and ground. The ground material (180 g) was extracted with Et₂O for 2 weeks. The crude extract (3.50 g) was chromatographed on Si gel using a n-hexane-EtOAc gradient. The first fraction (n-hexane) contained a mixture of sesquiterpene hydrocarbons (110 mg) in which α -elemene and β -elemene, γ cadinene and calamenene were detected by GC-MS. The above mixture was rechromatographed on Si gel using n-hexane to afford $(+)-\beta$ -elemene (1) (20 mg): $[\alpha]_D + 17.3^\circ$ (c, 1.20)(lit. + 15.1° [28]). The second fraction (n-hexane-EtOAc, 9:1) gave a mixture of carotenoids (30 mg). The third fraction (4:1) (1.720 g) was rechromatographed on Si gel using C₆H₆-EtOAc gradient to afford 6-[3-methyl-2-butenyl]-indole (720 mg) [11, 12], triglycerides (350 mg) and sterols (70 mg). The fourth fraction (1:1) gave fatty acids (530 mg).

A. pinguis Dum. (=R. pinguis) collected in Kochi prefecture in August 1978 and R. jackii Schiffn. in Kamikatsu-cho, Tokushima prefecture in May 1979 were treated in the same manner as R. multifida. Each crude extract (520 mg of A. pinguis and 2.330 g of R. jackii) was chromatographed on Si gel using the solvent system n-hexane-EtOAc. The first fraction (n-hexane) from A. pinguis (32 mg) contained at least six unidentified sesquiterpene hydrocarbons. The second fraction (n-hexane-EtOAc, 9:1) was a viscous fragrant oil which was rechromatographed on Si gel using C₆H₆ to afford carotenoids (10 mg), pinguisone (7) (80 mg) [14] and unidentified sesquiterpenes (20 mg). The third fraction (4:1) contained sterols (20 mg) and fourth fraction (1:1) contained fatty acids (45 mg). The first fraction (n-hexane) from R. jackii contained sesquiterpene hydrocarbons (220 mg) which were rechromatographed on Si gel impregnated with 5 % AgNO₃ using n-hexane as a solvent and then divided into two fractions: the first fraction was purified by preparative GLC to afford: (+)α-selinene (2) (25 mg), $[α]_D + 6.6^\circ$ (c, 1.10) (lit. +7.8° [29]); (-)- β -selinene (3) (18 mg), $[\alpha]_D - 32.5^\circ$ (c, 0.90) (lit. $+63^\circ$ [30]); (+)- α -barbatene (9) (23 mg), $[\alpha]_D + 44.7^\circ$ (c, 1.30) (lit. $+48^\circ$ [31]).

P. longispina Steph. collected in Gotaki, Tokushima prefecture in May 1979 was treated as described above. The crude extract (7.5 g) was chromatographed on Si gel using an n-hexane-EtOAc gradient. The first fraction (n-hexane) contained a mixture of paraffins and sesquiterpenes (350 mg) in which calamenene and cuparene (5) were detected by GC-MS as minor components. The second fraction (n-hexane-EtOAc, 19:1) (500 mg) was rechromatographed on Si gel using C_6H_6 to give a mixture of carotenoids (50 mg) and a mixture of sesquiterpenoids (240 mg). The third fraction (9:1) gave triglycerides (2.805 g). The fourth fraction (9:1) gave (-)-spathulenol (6) (500 mg). The fifth fraction gave fatty acids (1.700 g).

Acknowledgements—We wish to thank Dr. S. Hattori and Dr. M. Mizutani of Hattori Botanical Laboratory, Miyazaki, Japan, for their identification of the species and for useful suggestions.

REFERENCES

- Asakawa, Y., Tokunaga, N., Toyota, M., Takemoto, T. and Suire, C. (1979) J. Hattori Bot Lab. 45, 395.
- Asakawa, Y., Tokunaga, N., Toyota, M., Takemoto, T., Hattori, S., Mizutani, M. and Suire, C. (1979) J. Hattori Bot. Lab. 46, 67.
- Asakawa, Y., Hattori, S., Mizutani, M., Tokunga, N. and Takemoto, T. (1979) J. Hattori Bot. Lab. 46, 77.
- Asakawa, Y., Tokunaga, N., Takemoto, T., Hattori, S., Mizutani, M. and Suire, C. (1980) J. Hattori Bot. Lab. 47, 153
- Asakawa, Y., Suire, C., Toyota, M., Tokunaga, N., Takemoto, T., Hattori, S. and Mizutani, M. (1980) J. Hattori Bot. Lab. 48, 285.
- Asakawa, Y., Inoue, H., Toyota, M. and Takemoto, T. (1980) Phytochemistry 19, 2623.
- 7. Douin, R. (1958) C. R. Acad. Sci. Paris 246, 1248.
- 8. Das, V. S. R. and Rao, M. P. (1963) Nature 198, 970.
- 9. Das, V. S. R. and Rao, M. P. (1966) Current Sci. 35, 20.
- Das, V. S. R. and Rao, M. P. (1967) Indian J. Exp. Biol. 5, 193.
- Benesova, V., Samek, Z., Herout, V. and Sorm, F. (1969) Collect. Czech. Chem. Commun. 34, 1807.
- Benesova, V., Herout, V. and Sorm, F. (1969) Collect. Czech. Chem. Commun. 34, 1810.
- 13. Gorham, J. (1977) Phytochemistry 16, 249.

728 Y. Asakawa et al.

14. Benesova, V., Samek, Z., Herout, V. and Sorm, F. (1969) Collect. Czech. Chem. Commun. 34, 582.

- Andersen, N. H., Ohta, Y., Liu, C.-B., Kramer, M., Allison, K. and Huneck, S. (1977) Phytochemistry 16, 1727.
- Asakawa, Y. and Takemoto, T. (1978) Phytochemistry 17, 153.
- 17. Asakawa, Y., Toyota, M. and Takemoto, T. (1980) Phytochemistry 19, 1799.
- 18. Asakawa, Y. and Aratani, T. (1976) Bull. Soc. Chim. Fr. 1469.
- Asakawa, Y., Toyota, M., Uemoto, M. and Aratani, T. (1976) Phytochemistry 15, 1929.
- Asakawa, Y., Takemoto, T., Toyota, M. and Aratani, T. (1977) Tetrahedron Letters 1407.
- Asakawa, Y., Yamamura, A., Waki, T. and Takemoto, T. (1980) Phytochemistry 19, 603.
- Asakawa, Y., Toyota, M., Kano, M. and Takemoto, T. (1980) Phytochemistry 19, 2651.
- 23. Hopkins, B. J. and Perold, G. W. (1974) *J. Chem. Soc. Perkin Trans.* 1, 32.
- Matsuo, A., Nakayama, M., Maeda, T., Noda, Y. and Hayashi, S. (1975) Phytochemistry 14, 1037.

- Kazlauskas, R., Murphy, P. T., Quinn, R. J. and Wells, R. J. (1976) Aust. J. Chem. 29, 2533.
- Kalauskas, R., Murphy, P. T., Wells, R. J., Daly, J. J. and Oberhansli, W. E. (1977) Aust. J. Chem. 30, 2679.
- 27. Rose, A. F., Sims, J. J., Wing, R. M. and Wiger, G. M. (1978) Tetrahedron Letters 2533.
- Weinheimer, A. J., Middlebrook, R. E., Bledsoe, J. O., Jr., Marsico, W. E. and Karns, T. K. B. (1968) J. Chem. Soc. Chem. Commun. 384.
- Andersen, N. H., Shunk, B. and Costin, C. R. (1973) *Experientia* 29, 645.
- Ruzicka, L., Wind, A. H. and Koolhaas, D. R. (1931) Helv. Chim. Acta 14, 1132.
- Andersen, N. H., Bissonette, P., Liu, C.-B., Shunk, B., Ohta, Y., Tseng, C.-L. W., Moore, A. and Huneck, S. (1978) Phytochemistry 17, 1731.
- 32. Nishimura, K., Shinoda, N. and Hirose, Y. (1969) Tetrahedron Letters 3097.
- 33. Enzell, C. and Erdtman, H. (1968) Tetrahedron 4, 361.
- 34. Garbish, E. W., Jr. (1962) J. Org. Chem. 27, 4249.